

GLYCOSIDASE WEE-TABS

PRINCIPLE/DISCUSSION:

Some bacteria produce enzymes which hydrolyze various chromogenic substrates. Because the organism produces these enzymes during the growth process and the presence of the enzymes can be detected rapidly, more substrates can be used in a wide variety of applications which would normally require special growth conditions. KEY glycosidase tablets provide an easy to setup, inexpensive way to detect these enzymes.

Glycosidase: When bound to nitrophenol, the hydrolysis of the colorless aryl-substituted glycoside or phosphoester releases the nitrophenol base with its yellow color. The most common test using this principle is o-Nitrophenol- β -d-galactopyranoside (ONPG) used for enterics and *Neisseria* species.

ACTIVE INGREDIENTS:

The tablets contain approximately 0.05 mg. of one or more of the following substrates (see catalog #) in a Sodium Chloride and Dicalcium phosphate base:

- pNitrophenolNacetyl α dglucosaminide (NAG)
- oNitrophenol α darabinopyranoside (α ARA)
- pNitrophenol α lucopyranoside (α FUC))
- p-Nitrophenol- β -d-fucopyranoside (BFUC)
- oNitrophenol α dgalactopyranoside (α GAL)
- o-Nitrophenol- β -d-galactopyranoside (ONPG)
- pNitrophenol α dglucopyranoside (α GLU)
- p-Nitrophenol- β -d-glucopyranoside (BGLU)
- oNitrophenol α dmannopyranoside (α MAN)
- p-Nitrophenol phosphate (alkaline phosphatase) (PO4)
- o-Nitrophenol- β -d-xylopyranoside (BXYL)

MATERIAL SAFETY DATA:

None of the nitrophenol bases are known at this time to be hazardous. When used only as directed there is no hazard involved.

MATERIAL REQUIRED:

All tests require fresh 24 hour growth on solid media not available from KEY. Broth media are not acceptable. Consult a suitable manual for recommended media for the specimen. The following items are required but not provided:

- Inoculating loop
- Distilled water, pH 7.0-7.2
- Droppers

Each product is sold ready-to-use, 28 tubes per pack. The catalog numbers of the tablets are listed in the QUALITY CONTROL section.

SINGLE TEST SETUP:

- (1) Add 3-5 drops of distilled water to the test tube.
- (2) Inoculate heavily with a loopful of organism from a fresh pure 24 hour culture plate or slant. Mix with the loop until the organism is in suspension.
- (3) Go to step 3 below.

MULTIPLE TEST SETUP:

- 1) Harvest sufficient colonies into 1-2 ml of distilled water to make a suspension equal to #5 McFarland. This suspension may be stored aerobically for later use. (up to 5 days).
- 2) Add 3-5 drops of this solution to the tube containing the tablet, shaking to disintegrate the tablet.
- 3) Incubate all tests aerobically, uncovered, at 34-37C for 2 hours. Tests may be incubated up to 24 hours.

INTERPRETATION OF RESULTS:

No change at 2 hours is negative. The appearance of a yellow color at any time during the 2 hours is a positive glycosidase test. At 24 hours only very bright yellow is positive. PO4 tablets should only be considered positive at any time if very deep yellow. SEE **TIPS**.

TIPS:

Vortexing or shaking the test vigorously will enhance color development. A denser suspension will also produce brighter and faster reactions.

STORAGE:

Consult individual package label for storage instructions.

QUALITY CONTROL:

Each lot of tablets should be tested with known positive and negative organisms. Some suggested strains are listed. Dispose of all used material in a manner appropriate for biohazardous material.

Organism codes

01. <i>Arcanobacterium spp.</i>	02. <i>Bacteroides fragilis</i>
03. <i>Bacteroides ureolyticus</i>	04. <i>Corynebacterium spp.</i>
05. <i>Escherichia coli</i>	06. <i>Klebsiella pneumoniae</i>
07. <i>Bacteroides cacae</i>	08. <i>Prevotella denticola</i>
09. <i>Streptococcus intermedius</i>	10. <i>Streptococcus constellatus</i>

GLYCOSIDASE: POSITIVE NEGATIVE

K1046 α ARA	07.	08.
K1235 α FUC	02.	03.
K1230 BFUC	09.	10.
K1270 α GLU	02.	03.
K1275 BGLU	02.	03.
K1273 α GAL	02.	03.
K1442 α MAN	01.	04.
K1463 NAG	02.	03.
K1490 ONPG	02.	03.
K1045 PO4	02.	03.
K1692 BXYL	06.	05.

REFERENCES:

- (1) Manual of Clinical Microbiology, Fifth Edition, Chapter 36, Enterobacteriaceae
- (2) Kilian, M and Bulow, P. 1976. Rapid Diagnosis of Enterobacteriaceae, Acta path. microbio. Scan, Sect B, 84:245-251.
- (3) Wadsworth Anaerobic Bacteriology Manual, 5th Edition, 1993, Glucosidase tests, page 152.



KEY SCIENTIFIC PRODUCTS, INC
1113 EAST REYNOLDS STREET
STAMFORD, TEXAS 79553
VOICE 800-843-1539
FAX 888-440-4208
WWW.KEYSCIENTIFIC.COM

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